HYDROXYLATION OF DAMMARANE TYPE TRITERPENES WITH m-CHLOROPERBENZOIC ACID¹⁾

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Dammaran-20(S)-ol and 20(S)-hydroxydammaran-3 β -yl acetate were reacted with m-CPBA to give the corresponding 5 α -ol, 7 β -ol, 12 β -ol, 25-ol, 2 α -ol, and 5 α ,25-diol.

Oxidation reactions of natural products are very important from not only a synthetic but also a biological point of view. Much effort has been expended in simulating biological oxidations²⁾ and in developing synthetic methodologies.³⁾ Microbial oxidation,⁴⁾ oxidation using mammals,⁵⁾ dry ozonation,⁶⁾ and remote oxidation⁷⁾ have been given much attention and investigated extensively. None of these, however, is convenient as a synthetic method and ozonation can be dangerous.⁶⁾ m-Chloroperbenzoic acid was first used as an oxidizing agent to introduce a hydroxyl group at a bridgehead position of a bicyclic compound.⁸⁾ Since then, Takaishi <u>et al</u>.⁹⁾ have reported the application of m-CPBA to hydroxylation at bridgehead positions in various polycyclic compounds. We report here a simple method for the introduction of hydroxyl groups into triterpene skeletons and the preparation of a Ginseng sapogenin by use of m-CPBA.

Dammaran-20(S)-ol (1) was treated with m-CPBA (1.2 equiv.) in chloroform under reflux for 6 h. The usual work up and column chromatography on silica gel gave the 5 α -ol (2, 59 %),¹⁰⁾ 7 β -ol (3, 2.1 %),¹⁰⁾ 12 β -ol (4, 1.3 %),¹⁰⁾ 25-ol (5, 28 %),¹⁰⁾ 2 α -ol (6, 4.2 %),¹⁰⁾ and 5 α ,25-diol (7, 1.3 %)¹⁰⁾ as well as the starting material (1) recovered in 77 % yield. The ¹H NMR spectrum of 2 showed no peak between 3 and 5 ppm and in the ¹³C NMR spectrum thirty signals were observed, two of which were due to oxygen-bearing carbons [$\delta_{\rm C}$ 77.3 (C-5) and 75.4 (C-20)].¹¹⁾ The mass spectrum showed a molecular ion peak at m/z 446 and a characteristic peak at m/z 362 (Fig. 2). These facts suggested that one hydroxyl group had been introduced at the C-5 position. From careful assignment of the ¹³C NMR signals,^{11,12} it was concluded that 2 must be formulated as dammarane-5 α , 20(S)-diol (2).

In the ¹H NMR spectrum of 3, a double doublet centred at δ 3.80 (1H, J=11 and 5 Hz) was observed and the ^{I3}C NMR spectrum indicated the presence of thirty carbons, two of which bore oxygen functions [δ_{c} 75.3 (C-7) and 75.4 (C-20)].¹¹



From the coupling pattern of the methine proton, the hydroxy group could be assigned to the 1 β , 7 β , or 15 α position. Of the five quaternary carbon atoms $[\delta_{\rm C}$ 75.4 (C-20), 49.7 (C-14), 46.2 (C-8), 37.3 (C-10), and 33.2 (C-4)]¹¹⁾ of 3, four of them had similar chemical shifts to those of dammaran-20(S)-ol (1)[$\delta_{\rm C}$ 75.3 (C-20), 50.4 (C-14), 40.6 (C-8), 37.5 (C-10), and 33.4 (C-4)].¹²⁾ Namely the carbon signal at $\delta_{\rm C}$ 40.6 assigned to C-8 has undergone a downfield shift of 5.6 ppm,indicating that 7 β was the most probable position for the hydroxyl group. The secondary alcohol (3) was oxidized to the corresponding ketone (8)¹⁰ by Jones' reagent. Methylene protons were observed at δ 2.50 (1H, t, J=15 Hz) and 2.20 (1H, dd, J=15 and 2.5 Hz) in the ¹H NMR spectrum of 8. From these results the secondary alcohol was determined to be dammarane-7 β ,20(S)-diol (3).

The third product (<u>4</u>) was a secondary alcohol from its ¹H NMR spectrum [δ 3.60 (1H, td, J=11 and 5 Hz)] and ¹³C NMR spectrum [$\delta_{\rm C}$ 74.4 (C-20) and 71.2 (C-12)].¹¹⁾ The coupling pattern indicated that the position of the hydroxyl group was 6α , 11α , 12β , or 16 β . Comparison of the ¹³C chemical shifts with those of known dammarane triterpenes¹²) implied a 12 β -ol structure. Jones' oxidation of <u>4</u> gave ketone (<u>9</u>)¹⁰ whose ¹H NMR spectrum showed a doublet at δ 2.85 (1H, J=10 Hz, H-13 β), a doublet of doublets at δ 2.30 (1H, J=14 and 5 Hz, H-11 α), and a triplet at δ 2.19 (1H, J=14 Hz, H-11 β). From these facts the third product was concluded to be dammarane-12 β ,20(S)-diol (<u>4</u>).

The fourth product (5) was very easily assigned as dammarane-20(S),25-diol (5) from its mass spectrum [m/z 410 (M-H₂OX2), 191, 145, 127 (base), and 59]

and ¹H NMR spectrum (eight singlet methyls were observed at δ 1.23, 1.23, 1.14, 0.96, 0.88, 0.85, 0.84, and 0.81).

A triplet of triplets observed at δ 3.89 (1H, J=11 and 4.5 Hz) in the ¹H NMR spectrum of the penultimate product (<u>6</u>) indicated the presence of HO an equatorial secondary hydroxyl group with a methylene group on each side. The same pattern was observed in the ¹H NMR spectrum [δ 5.06 (1H, tt, J=10 and 6 Hz)] of its acetate (<u>10</u>).¹⁰ The position of



observed in the ¹H NMR spectrum [δ 5.06 (1H, tt, Fig. 2 J=10 and 6 Hz)] of its acetate (<u>10</u>).¹⁰ The position of substitution is limited only to 2 α (equatorial). The corresponding ketone (<u>11</u>)¹⁰ obtained by Jones' oxidation showed four protons α to the carbonyl group [δ 2.39 (1H, dd, J=13 and 2 Hz), 2.28 (1H, d, J=13 Hz), 2.15 (1H, dd, J=13 and 2 Hz), and 1.98 (1H, d, J= 13 Hz)]. These data were fully consistent with structure <u>6</u>, dammarane-2 α ,20(S)diol.

The most polar product (7) showed eight singlet methyls in its ¹H NMR spectrum and fragment peaks at m/z 444 (M-H₂O), 378 (Fig. 2), 127, and 59 in its mass spectrum, suggesting the presence of a 25-hydroxyl group. In the ¹³C NMR spectrum thirty signals were detected, three of which corresponded to oxygenated carbons [δ_c 77.4 (C-5), 75.4 (C-20), and 71.1 (C-25)].¹¹) Comparison of the spectral data with those of 2 and 5 led directly to structure 7.

We next carried out a similar reaction of 20(S)-hydroxydammaran-3 β -yl acetate (<u>12</u>) with <u>m</u>-CPBA in chloroform. Four products [<u>13</u> (m/z 486, 426, 362, and 344; δ 5.12 (lH, dd, J=ll and 6 Hz) and 2.03 (3H, s); δ_{c} [170.9 (CO), 78.9 (C-5), 77.2 (C-3), and 75.4 (C-20)), <u>14</u> (m/z 486, 433, 419, 108, and 95; δ 4.48 (1H, dd, J=ll and 6 Hz), 3.79 (lH, dd, J=ll and 4 Hz), and 2.05 (3H, s); δ_{c} 171.0 (CO), 80.6 (C-3), 75.4 (C-20), and 75.1 (C-7)), <u>15</u> (m/z 468, 419, 341, and 108; δ 4.48 (1H, dd, J=ll and 6 Hz), 3.60 (lH, td, J=l0 and 5 Hz), and 2.05 (3H, s); δ_{c} 171.0 (CO), 80.8 (C-3), 74.5 (C-20), and 70.9 (C-12)), and <u>16</u> (m/z 468, 189, 145, 136, 127, and 109; δ 4.48 (1H, dd, J=l1 and 6 Hz) and 2.05 (3H, s); δ_{c} 171.0 (CO), 81.0 (C-3), 75.4 (C-20), and 71.0 (C-25))]^{10,11} were obtained in yields¹⁰ of 16, 1.6, 1.6, and 40 %, respectively, after column chromatography on silica gel (<u>12</u> was recovered in 91 % yield) and the structures were determined by comparing the spectral data with those of 2 - 7.

The spectral data of the third product (<u>15</u>) were completely identical with those of authentic 12 β ,20(S)-dihydroxydammaran-3 β -yl acetate (<u>15</u>).¹⁴) As this is an acetate of a typical sapogenin of Ginseng, this constitutes a one step synthesis of such types from a 12-deoxy compound. Tanaka and his group¹⁵) reported the first successful application to compounds other than steroids of the remote oxidation initially explored by Breslow and his co-workers.⁷) Photolysis of 20(S)-hydroxydammaran-3 α -yl p-nitrophenylacetate in Bu^tOH gave dammarane-3 α , 12 α ,20(S)-triol in 13 % yield after hydrolysis.¹⁵) Although the yield of <u>15</u> was not so good as compared to that reported, ¹⁵) our method is quite simple and easy to carry out.

The hydroxylation reaction reported here can be explained as an attack on unactivated carbon atoms. Although the mechanism is unknown, this method is quite useful for introduction of a functional group at unactivated carbon atoms. Further applications to sesquiterpenes, diterpenes, and other classes of triterpenes are under way.

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References and Notes

- 1) Part 2: Hydroxylation of Natural Products with <u>m-Chloroperbenzoic Acid</u>.
- 2) e.g. J. T. Groves and T. E. Nemo, J. Am. Chem. Soc., 105, 6243 (1983).
- <u>e.g.</u> J. Fuhrhop and G. Penzlin, "Organic Synthesis--Concepts, Methods, Starting Materials", Verlag Chemie, Weinheim (1983).
- 4) e.g. K. Takeda, Pure & Appl. Chem., 21, 181 (1970).
- 5) <u>e.g</u>. T. Ishida, Y. Asakawa, and T. Takemoto, J. Pharm. Sci., <u>71</u>, 965 (1982).
- 6) e.g. Y. Mazur, Pure & Appl. Chem., 41, 145 (1975).
- 7) e.g. R. Breslow, Chem. Soc. Rev., 1, 553 (1972).
- 8) W. Müller and H. -J. Schneider, Angew. Chem., Int. Ed. Engl., 18, 407 (1979).
- 9) N. Takaishi, Y. Fujikura, and Y. Inamoto, Synthesis, 1983, 293.
- 10) Satisfactory spectral data were obtained for all new compounds. ¹H and ¹³C NMR spectra were measured on a JEOL GX-400 (400 MHz for ¹H and 100 MHz for ¹³C) spectrometer in CDCl₃. Yields correspond to isolated yields based on consumed starting material.
- 11) Complete assignments of ¹³C NMR signals will be published elsewhere.
- 12) M. Tori, T. Tsuyuki, and T. Takahashi, Bull. Chem. Soc. Jpn., 50, 3349(1977).
- 13) J. Asakawa, R. Kasai, K. Yamasaki, and O. Tanaka, Tetrahedron, <u>33</u>, 1935 (1977).
- 14) Thanks are due to Prof. O. Tanaka, Hiroshima University, for kindly sending the sample of dammar-24-ene-3β,12β,20(S)-triol.
- 15) R. Kasai, K. Shinzo, O. Tanaka, and K. Kawai, Chem. Pharm. Bull., <u>25</u>, 1213 (1974).

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